SYNTHESIS OF 2-BROMO-L-TRYPTOPHAN AND 2-CHLORO-L-TRYPTOPHAN

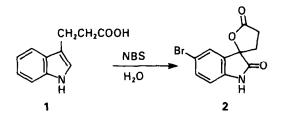
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<u>Abstract</u>: Free-radical halogenation of protected L-tryptophan with N-bromosuccinimide or N-chlorosuccinimide leads to the corresponding 2-halo derivative in high yield; enzymatic removal of the blocking groups provides the new amino acid analogues.

A wide variety of tryptophans with substituents in the benzenoid portion of the indole ring system have been prepared by chemical synthesis¹ and by biosynthesis.² In contrast, only a few analogs are known with substituents at C-2: methyl,³ hydroxy,^{4,5} thio,^{5b} thioether,⁶ and carboxy.⁷ As a consequence of the discovery, in this laboratory, of the widespread biological activities of 2-fluorohistidine,⁸ we became interested in 2-halotryptophans as potential irreversible inhibitors of tryptophan-utilizing mammalian enzymes—particularly indoleamine dioxygenase, which has been reported to be induced by interferon.⁹

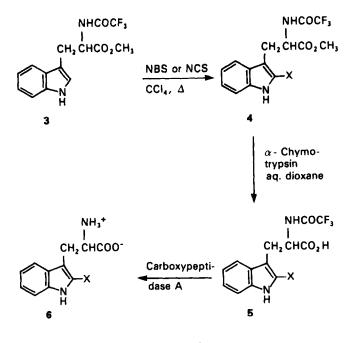
In protic solvents, halogenation of simple 3-alkylindoles leads to oxindoles with or without an additional function at C-3.¹⁰ Intramolecular participation has also been observed, indole-3-propionic acid (1) leading to a spirolactone (2);¹¹ analogous products



have been obtained with N-acyltryptophan or its peptides.¹² Spirolactone formation does not occur with N-acetyltryptophan esters but, under certain conditions, the α -nitrogen atom can become involved in cyclization to 2,3-dihydropyrrolo[2,3-b]indoles.¹³

Under aprotic or radical-promoting conditons, halogenation of 3-alkylindoles leads to the 2-halo derivatives quite cleanly.¹⁴ Esters of indole-3-acetic acid have been similarly halogenated in CCl_4 as solvent¹⁵ and C-2 halogenation of lysergic acid derivatives was effected in dioxane.¹⁶

On the basis of these precedents, we have now found that α -N-trifluoroacetyl-Ltryptophan methyl ester (3) is converted into its 2-bromo derivative (4a) in 83% isolated yield, following reaction with NBS/benzoyl peroxide in refluxing CCl₁. The NMR spectrum of the bromination product clearly establishes the position of substitution as C-2, since the sharp singlet due to H-2 in 3 (δ 6.94) is completely absent in the product while the spectrum of aromatic protons is unchanged. Conversion of <u>4a</u> to the amino acid (<u>6a</u>) by acid hydrolysis was excluded because of the well-documented lability of 2-haloindoles in aqueous acid.¹⁰ On the other hand, the observations of alkaline stability of 2-haloindoles¹⁰ did not extend to <u>4a</u> or <u>6a</u>; following exposure of <u>4a</u> to 1 N NaOH, TLC revealed a multiplicity of ninhydrin-positive products. We then considered enzymatic methods and found that α -chymotrypsin, by virtue of its rather broad specificity,¹⁸ hydrolyzed <u>4a</u> to <u>5a</u> (mp 148-150 °C) in 60% yield at neutral pH.¹⁹ Hydrolysis of the trifluoroacetyl group was then performed by use of carboxypeptidase A²⁰ and the free amino acid (<u>6a</u>) was isolated in 70% yield.



a, X=Br; b, X=Cl

Compound <u>6a</u> reacts readily with $AgNO_3$ and is converted into oxindolylalanine under mild conditions (25 °C, pH 1, 2-3 h).

An attempt to brominate α -N-acetyltryptophan methyl ester with NBS in CCl₄ did not result in an isolable yield of the expected product; however, bromination of methyl indole-3-propionate gave methyl 2-bromoindole-3-propionate in 72% yield.

Chlorination of 3 with N-chlorosuccinimide in CCl_4 provided 4b, mp 150-152 °C, in 55% yield.²¹ As with the bromo derivative, chymotrypsin was used to hydrolyze the ester to 5b, and carboxypeptidase A removed the trifluoroacetyl group to give 2-chlorotryptophan (<u>6b</u>).

The UV spectrum of 2-bromotryptophan is very similar to that of tryptophan $[\lambda_{max}(H_20) \ 218 \ (\log \ \epsilon \ 4.48), \ 279 \ (\log \ \epsilon \ 3.84) \ and \ 288 \ nm \ (\log \ \epsilon \ 3.75)], \ although the extinction coefficient for <u>6a</u> is ca. 50% greater than for tryptophan in the 280 \ nm region. The UV spectrum of 2-chlorotryptophan shows <math>\lambda_{max}$ at 273 rather than at 280 nm; 2-chloro-

skatole¹⁴ and 2-chloroindole-3-acetic acid¹⁵ have also been reported to exhibit λ_{max} at 273-274 nm. While aqueous solutions of tryptophan are highly fluorescent, exhibiting strong emission at 355 nm upon excitation at 280-300 nm, neither 2-chloro- nor 2-bromo-tryptophan exhibits any significant fluorescence under these conditions.

Both in aqueous solution and in the solid state, $\underline{4a}$ and $\underline{4b}$ have given no evidence of instability. On the other hand, dried residues of the noncrystalline amino acids ($\underline{6a}$ and $\underline{6b}$) darken gradually at ambient temperature but are stable at -20 °C; at neutral pH, aqueous solutions of the amino acids are stable for at least 48 h at room temperature.

Hydrolysis of 2-haloindoles is subject to catalysis in 1-3 N acid, with k showing a linear dependence of acid activity (h_0) . The 2-chloro compounds are 1.5-2 times as reactive as the corresponding 2-bromo compounds. Measurement of hydrolysis kinetics over a wide pH range revealed, however, that compounds with a free carboxyl group in the side chain (5a, 6a, etc.) are also subject to <u>intramolecular</u> acid catalysis (Table I), that intramolecular catalysis is considerably more effective than bimolecular catalysis, and that the relative efficiency of the intramolecular pathway increases rapidly with pH. Since it is quite likely that the function of the catalyst acid is to convert the stable 2-haloindole into an unstable 2-haloindolenine by protonation of C-3, ²² we would hope that such activation by proton transfer may also occur within a biological tryptophan-recognition site and result in affinity labeling. Further studies on the chemical and biological properties of these novel amino acids are in progress.

Table I. Half-1	ives for Hydrolysis	at 25 °C (hrs)
Compd	1 N HC104	рН 2
2-Br-skatole	19	1920
<u>4a</u>	~19	~1900
<u>5a</u>	0.7	1.6
6 <u>a</u>	0.6	1.4

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- 17. The compound crystallized from ethyl acetate-petroleum ether, mp 154.5-155.5 °C; it was fully characterized by MS (CI, NH₃) [m/z 410 (100), 412 (95), M + 18; 393 (18), 395 (18), M + 1], NMR (220 MHz, $CDCl_3/TMS$) [δ 8.23 (s, 1, indole NH), 7.1-7.5 (m's, 4, aryl H's), 6.93 (bs, 1, α -NH), 4.92 (m, 1, α -CH), 3.73 (s, 3, OCH_3), and 3.35 (d, 2, β -CH₂)], and elemental analysis [Calcd. for $C_{14}H_{12}BrF_3N_2O_3$: C, 42.77; H, 3.08; N, 7.13; Br, 20.32. Found: C, 43.00; H, 3.15; N, 7.03; Br, 20.36].
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(Received in USA 26 September 1983)